Lundquist Institute for Biomedical Innovation
at Harbor-UCLA Medical Center

Institutional Biosafety Committee
Meeting Minutes
09/09/2025
Zoom Virtual Conference

MEMBERS PRESENT	MEMBERS ABSENT
David Applebaum, M.S.	Fawzia Bardag-Gorce, Ph.D.
Helen Chun, Ph.D.	Fang Wang, Ph.D.
Rami Doueiri, Ph.D.	
Scott Filler, M.D.	
Adrienne Zweifel, Ph.D.	

STAFF PRESENT	STAFF ABSENT
Rosa Harmon, CPIA	Elizabeth Burrola, CIP
Annie Hilo	Rosemary Madnick, MBA

1. CALL TO ORDER

The meeting was called to order by Scott Filler, M.D. at 3:01 PM.

2. MEETING MINUTES

The minutes of the August 12, 2025 meeting were presented.

A motion was made and seconded to APPROVE the minutes.

Vote: For - 5, Opposed - 0, Absent – 0, Abstained - 0, Recused – 0

3. BUA REVIEW

A. AMENDMENTS

IBC #: IBC 31789-01
PI: Priya Uppuluri, Ph.D.
iRIS Ref #: 062881

Summary: Experiments involve the use of opportunistic fungal pathogens, including *Candida albicans*, *Candida auris*, other *Candida* species, *Aspergillus fumigatus*, *Rhizopus* species, and dermatophytes such as *Trichophyton* species. The animal host will be infected with fungal infections using different routes of infection for evaluating virulence or response to drugs.

Inserts: TBD

Vectors: NA

Pathogens: *Aspergillus fumigatus, Rhizopus delemar, Candida species, Trichophyton mentagrophytes*

Risk Assessment: Risk Group 2

Containment: BSL-2

Training: Training has been verified by the Research Compliance Office Staff and EHS. It was noted upon review of the BUA that updated training dates for the study personnel is needed or the individual must undergo the required refresher training.

NIH Guidelines: NA

Conflicts: None.

Motion: A motion was made and seconded to REQUIRE MODIFICATIONS TO SECURE APPROVAL of the BUA.

Modifications Required to BUA:

1. **Section 4.0 - Background Information**
   - For Section 4.4, the PI is to ensure personnel has updated Biosafety training and revise to include the most current training date.

2. **Section 5.0 - Description of Procedure(s)/Research**
   - Lay Description – The PI is to update the existing text with "Mice will be infected with fungal *pathogens* using different routes of inoculation for evaluating virulence or response to drugs."
   - Overall Goal – The PI is to explicitly state which species will be genetically modified and describe the types of genetic modifications made. Overall, the PI is to define mutant library and explain how it was generated.
   - Experimental Procedures - The PI is to:
     - clarify what other Candida species are in the lab's possession.
     - Describe how fungal spores will be collected in the lab (inoculum for animal experiments).
     - Describe how each route of inoculation will be performed (intratracheal, intranasal, inhalation, intravaginal, etc).
     - Briefly explain how skin colonization, per Candida species entry in Section 8.0 (Entry #3), is done.
     - Provide examples of the fomites/abiotic surfaces that will be used in this work.
     - Provide details on how agents will be applied to fomites/abiotic surfaces.
     - Clarify what is meant by a "dedicated animal biosafety cabinets." Is this a BSC in the animal procedure space, or a cage change station or other engineering control?
     - Confirm that eye protection will be worn "at all times," even when working in a BSC.
     - Explicitly state whether respiratory protection (N95 or PAPR) will be used for all agents, or just agents whose route of inoculation includes inhalation or an aerosol-generating procedure.

The N95 respirators might not be necessary as many of the fungi are not known to spread by aerosols. If the study team would like to use N95 respirators, the PI is to list the individuals who will use them and they will be required to be trained, fit tested and medically evaluated in accordance with TLI's Respiratory Protection Program. The PI is to contact the EH&S department to schedule the training, fit testing and medical evaluations for those individuals.

- Discuss any sampling performed on infected animals and whether this sampling is done on awake, but restrained, mice or if the mice will be anesthetized.
- Explicitly state whether tissues, organs, etc. will be harvested and processed. If yes, the PI is to describe how this is performed and what the lab is looking for.

- Hazard Assessment – The PI is to:
  - Instead of stating "BSL2+," the PI is to explicitly state what enhancements will be employed for work involving C. auris.
  - A member of the PI's team mentioned during the lab audit that the lab uses 50% household bleach for dermatophyte work citing that this species was resistant to 10% bleach. The PI is to update this section to include this information, or provide training to lab staff regarding the effective concentration of disinfectant to be used with this pathogen.
  - Explicitly state which disinfectant is mainly used in the lab for which agents.
  - Replace "waste materials will be autoclaved prior to disposal" since the lab is not autoclaving waste before disposal. Consider "waste materials will be disposed of via the medical waste stream."
  - Remove personnel clothing as a potential vector for transmission as the PPE in use should be selected to protect staff clothing from contamination. If street clothes are known to be contaminated, then lab staff would not be allowed to take them home to clean. If staff change out of their street clothes into scrubs when working with these pathogens, the PI is to explicitly state this.

- Emergency Procedures - the PI is to explicitly state their use of 50% household bleach in the event of a spill.

3. **Section 8.0 - Infectious Agents/Toxins (Excluding Viral Vectors)**
   - Candida species - Other than C. auris and C. albicans, the PI is to clarify what other Candida species are used and create individual entries for each of the Candida species.
   - Trichophyton mentagrophytes - The PI is to provide a reference showing that 70% ethanol is effective against this agent.

4. **Section 10.0 - Encoded Recombinant or Synthetic DNA Sequences**
   - The question "Will the research use encoded recombinant or synthetic DNA sequences" is marked no, but the lab referenced mutant libraries in Section 5.0 of the BUA. The PI is to clearly state how the mutant libraries are generated. The answer should be marked "Yes" if any of these agents have been genetically modified, even if that modification was not performed in their lab (i.e., performed elsewhere). If there are any assays that involve human cell lines, those should also be included.

5. **Section 12.0 - Research with Eukaryotic Hosts (Including Fungi)**

- Since study team clarified the Trichophyton species will not be the host, the PI is to revise the response in Section 12.1 of the BUA and select "No".

Vote: For - 4, Opposed - 0, Absent – 1 (R. Doueiri), Abstained – 0, Recused – 0

B. CONTINUATIONS

IBC #: IBC 2025-U-Lenti OK
PI: Omid Khorram, M.D.
iRIS Ref #: 062775

Summary:

#32133: Freshly isolated leiomyoma cells will be transduced by lentivirus for knockdown of TDO2. For lentivirus generation and cell transduction, 293T cells will be transfected with lentivirus vectors (pCLucIPZ, a self-inactivating vector) carrying TDO2-targeting shRNAs along with package plasmids (pMD2.G, pRSV-Rev and pMDLg/pRRE) and supernatants will be collected. Primary leiomyoma cells will be transduced by the concentrated lentiviral suspension, and grafted under the kidney capsule of female anesthetized mice.

#31752: Freshly isolated leiomyoma cells from hysterectomy specimens will be transduced by lentivirus for knockdown of MIAT and/or XIST. Lentivirus control vector (pCLucIPZ) will be inserted with MIAT or XIST-targeting shRNAs. For lentivirus generation and cell transduction, briefly, 293T cells will be transfected with lentivirus vectors along with package plasmids (pMD2.G, pRSV-Rev and pMDLg/pRRE) and supernatants will be collected. Primary leiomyoma cells will be transduced with lentivirus and selected by culturing with puromycin for 3 days and then prepared into cell pellets. After overnight floating culture in complete medium, cells will be grafted onto kidney capsule of female anesthetized mice as follows: 1. Empty vector (pCLucIPZ); 2. MIAT knockdown; 3. XIST knockdown; 4. Both MIAT and XIST knockdown. After 8 weeks the animals will be sacrificed and cell cluster excised, measured and subjected to proposed experiments.

Primary leiomyoma material will be obtained directly from the hospital operating room under an IRB-approved tissue collection protocol.

Inserts: shTDO2, shXIST, shMIAT, Luciferase, puromycin-N-acetyltransferase (puromycin resistance gene)

Vectors: pRSV-Rev, pMD2.G, pCLucIPZ, pMDLg/pRRE

Pathogens: NA

Risk Assessment: Risk Group 2

Containment: BSL-2

Training: Training has been verified by the Research Compliance Office Staff and EHS.

NIH Guidelines: III-D, III-F

Conflicts: None.

Motion: A motion was made and seconded to REQUIRE MODIFICATIONS TO SECURE APPROVAL of the BUA.

Modifications Required to BUA:

1. **Section 5.0 - Description of Experiment(s)/Research**
   - Experimental Procedures - For #32133, the PI is to state how the cell cluster will be measured.
   - The PI is to clarify the "proposed experiments."

As previously requested:
   - Experimental Procedures – The PI is to provide more information on how harvested materials are processed and indicate the type of "further analysis."
   - Containment Conditions – The PI is to explicitly state that materials will be disposed of as medical waste.

2. **Section 9.0 - Vectors for Recombinant or Synthetic DNA**
   - The PI is to add the lentivirus packaging plasmids back in this section since these are used in the HEK cells in order to package the lentivirus.

3. **Section 11.0 - Research with Whole Plants or Animals as Hosts**
   - The PI is to revise the answer to this question to "Yes" since the lab is genetically modifying cell lines using lentivirus, then using those cells in an animal model.

Vote: For - 5, Opposed - 0, Absent - 0, Abstained – 0, Recused – 0

4. OTHER BUSINESS

a. Safety Committee Report – Accidents/Spills
Mr. Applebaum stated no incidents were reported.

b. Lab Coat Guidance - update
Mr. Applebaum reported the lab coats have been ordered by Purchasing.

Door Signage - update
Dr. Zweifel inquired about door signage and Mr. Applebaum confirmed that door signs have been posted. In addition, a sign is posted in the elevator area on the 2nd floor indicating the types of PPE required to work in that area.

c. News from the NIH Office of Science Policy
"NIH Launches Comprehensive Effort to Modernize Biosafety Framework" dated 9/9/2025
The recent announcement from the NIH regarding the launch of a new biosafety modernization initiative to strengthen biosafety policies, practices and oversight was shared

with the Committee. The Committee briefly discussed how these future changes may impact local research.

d. Self-Assessment Tool: Surveillance, Emergency Planning, and Response section
The Committee reviewed items 46 to 53 in the Surveillance, Emergency Planning, and Response section, and the discussion is to be noted within the document maintained on file in the Research Compliance Office.

With no further business, the meeting adjourned at 3:46 PM

Respectfully submitted,

Signed by:
Scott Filler
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Scott Filler, M.D.
Chair, Institutional Biosafety Committee

cc: Research Committee Agenda